

# Citrus Huanglongbing Diagnosis Based on Molecular Detection of Associated Liberibacter Species

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# Major management strategy for HLB control

- To remove infected host plants
- To reduce suspect psyllid populations

Methods to identify the disease or its associated bacteria, currently three known species, '**Ca. Liberibacter asiaticus**', '**Ca. L. africanus**' and '**Ca. L. americanus**'

plants  
and  
psyllids



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# Yellow shoots (China) for HLB finding



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# Leaf mottle (Philippines) for HLB finding



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# Vein degeneration (Indonesia) for HLB finding



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# Dieback (India) for HLB finding



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# Fruit symptoms (S. Africa) for HLB finding



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# Traditional methods to detect the HLB-associated bacteria

- **Electron microscopy**: Lafleche & Bové, 1970.
- **ELISA**: Garnier et al., 1987.
- **Iodine reaction (IR)**: Schneider H, 1968.
- **Fluorescent substance**: Schwarz, 1968
- **Biological indexing**: Schwarz, 1968.



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# Nucleic acid-based non-PCR methods to detect the HLB-associated bacteria

- **Dot-blot hybridization**: low sensitivity as electron microscopy (Villechanoux *et al.*, 1992).
- **Loop-mediated isothermal amplification (LAMP)**: useful for labs without a PCR machine but vulnerable to contaminations (Okuda *et al.* 2005).
- **Cycleleave isothermal and chimeric primer-initiated amplification of nucleic acids with probe technology (ICAN)**: useful for screening labs without PCR machines (Orasaki *et al.*, 2008).



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# PCR methods for detection of HLB-associated bacteria



Kary B. Bullis  
California USA



Michael Smith  
Vancouver Canada

Since its first introduction in 1983 by the two Nobel prize winners, PCR has become a powerful technique for detection and identification of plant pathogens



Dr. Bové's lab developed first PCR primers for detection and identification of the HLB-associated *Liberibacter* species (Jagoueix *et al.*, 1996).

The main objective of the presentation is to discuss how to use PCR methods for detection, identification and quantification of the HLB bacteria

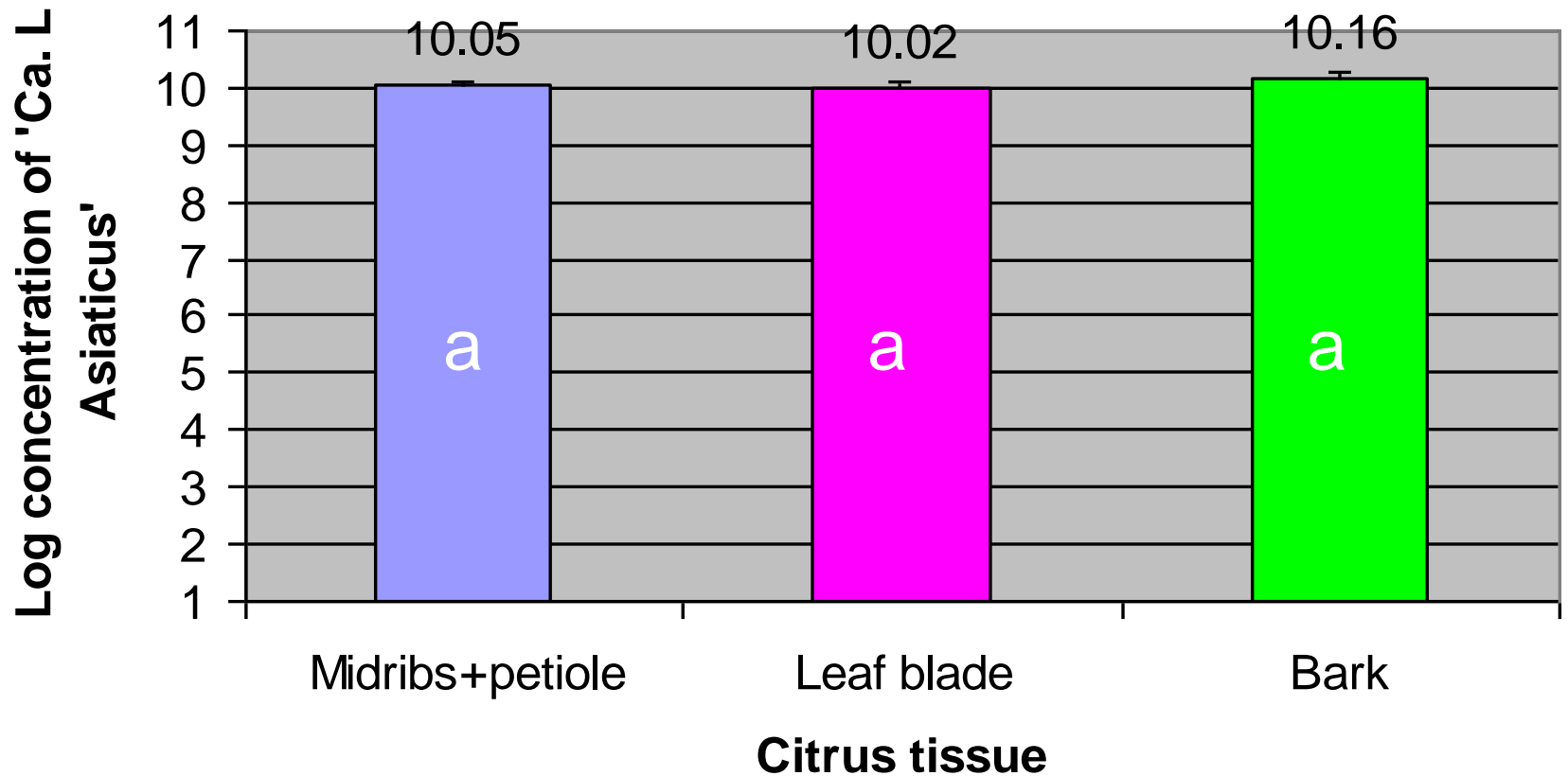


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# Quantification of 'Ca. L. asiaticus' in HLB-affected citrus tissues



Li *et al.*, 2009. *Phytopathology* 99:139-144



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## Sampling HLB suspect trees

1 to 4 branches (twigs) with symptomatic leaves or fruit from each tree

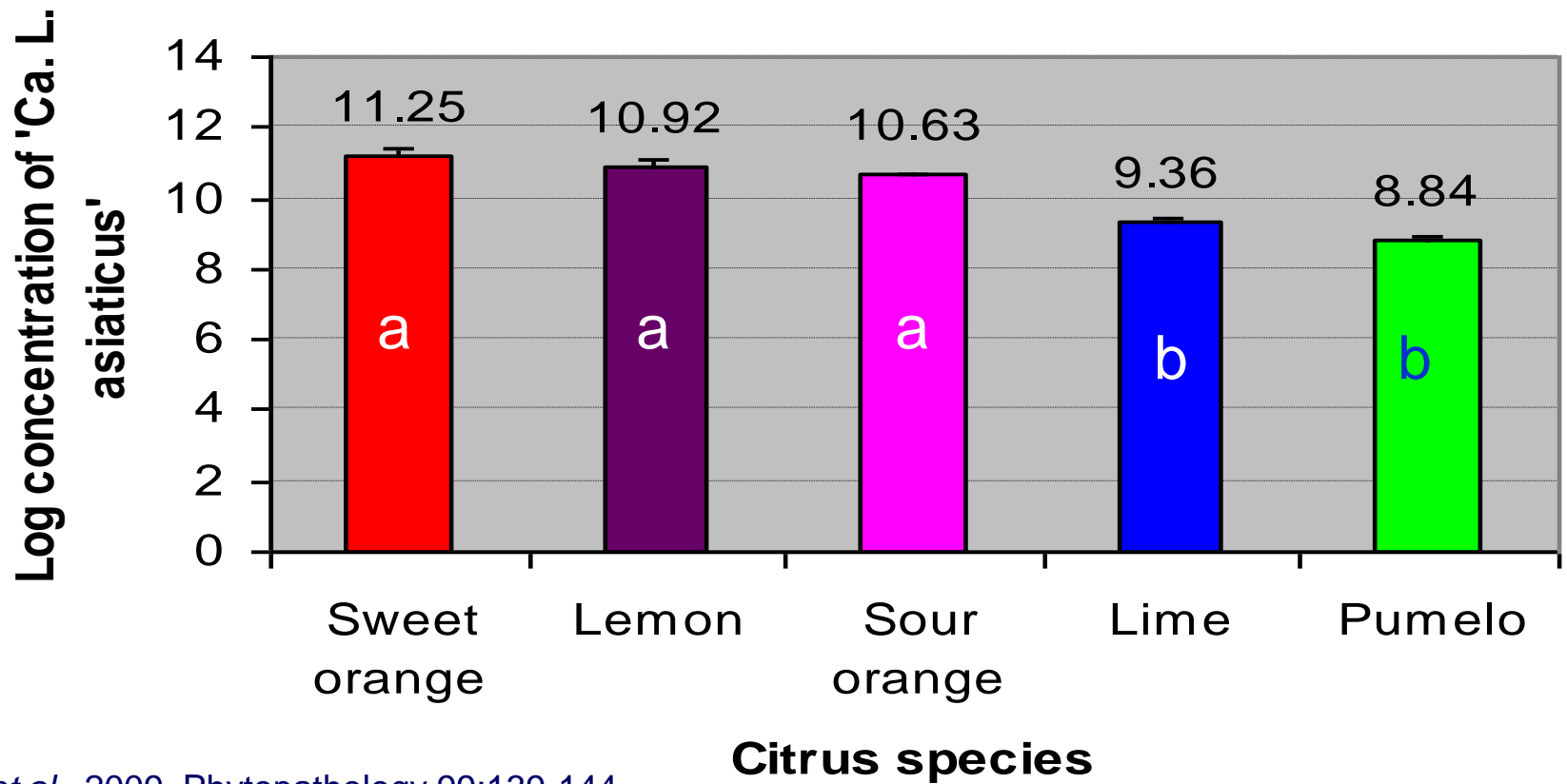


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# Quantification of 'Ca. L. asiaticus' in midribs and petioles of HLB-affected citrus species



Li *et al.*, 2009. *Phytopathology* 99:139-144

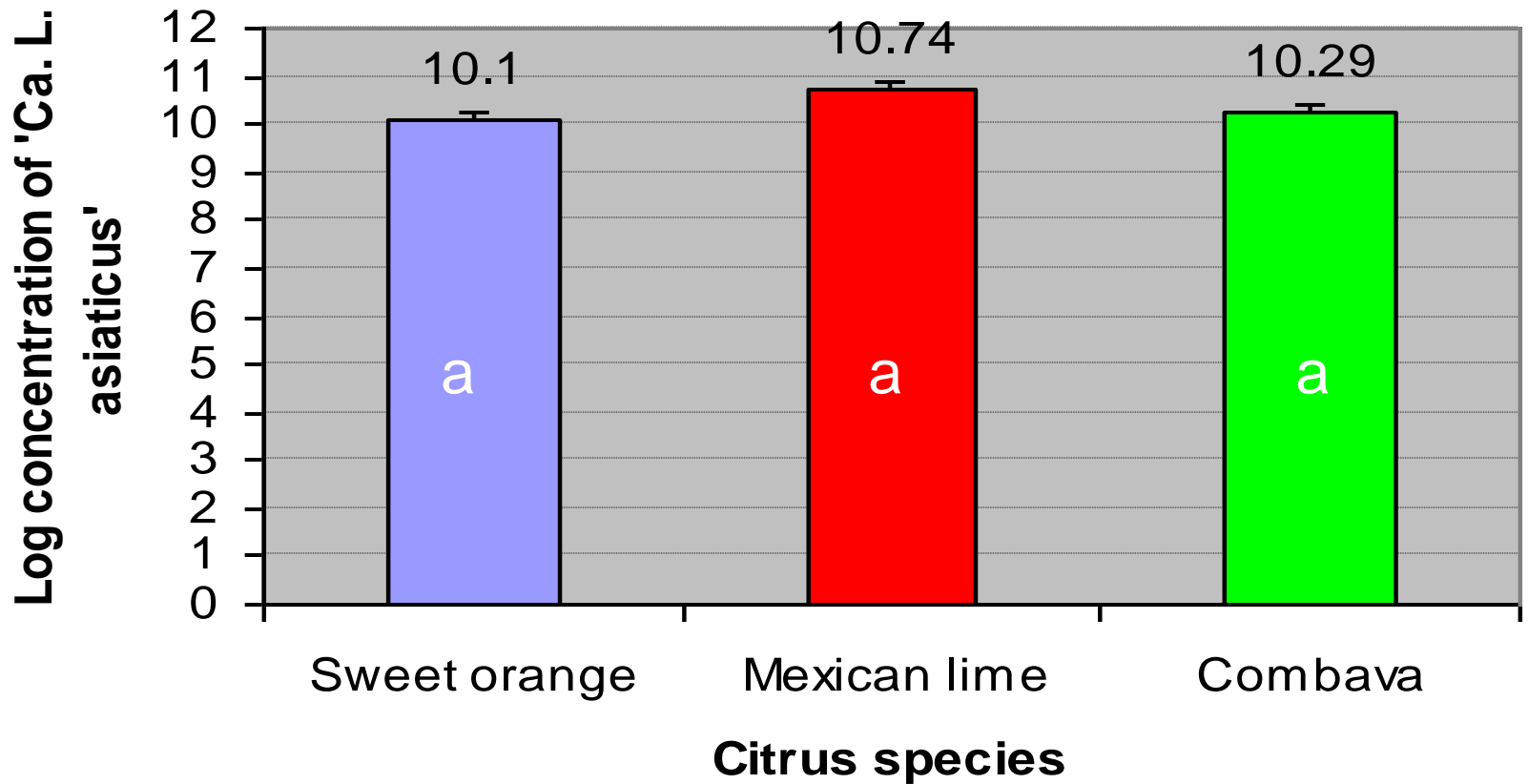


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# Quantification of 'Ca. L. asiaticus' in root bark of HLB-affected citrus species



Li *et al.*, 2009. *Phytopathology* 99:139-144

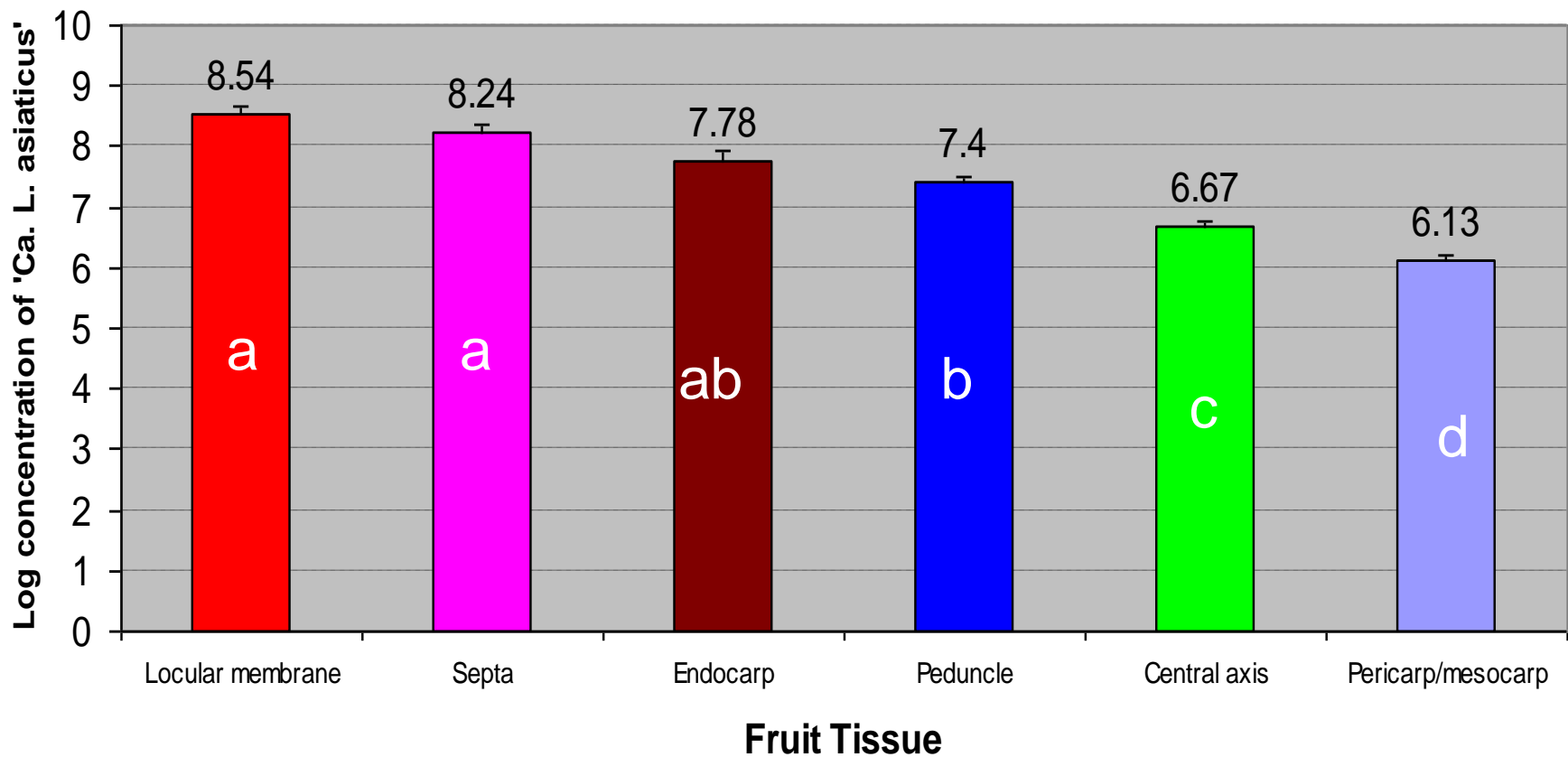


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# Quantification of 'Ca. L. asiaticus' in fruit tissues of HLB-affected citrus species



Li *et al.*, 2009. *Phytopathology* 99:139-144

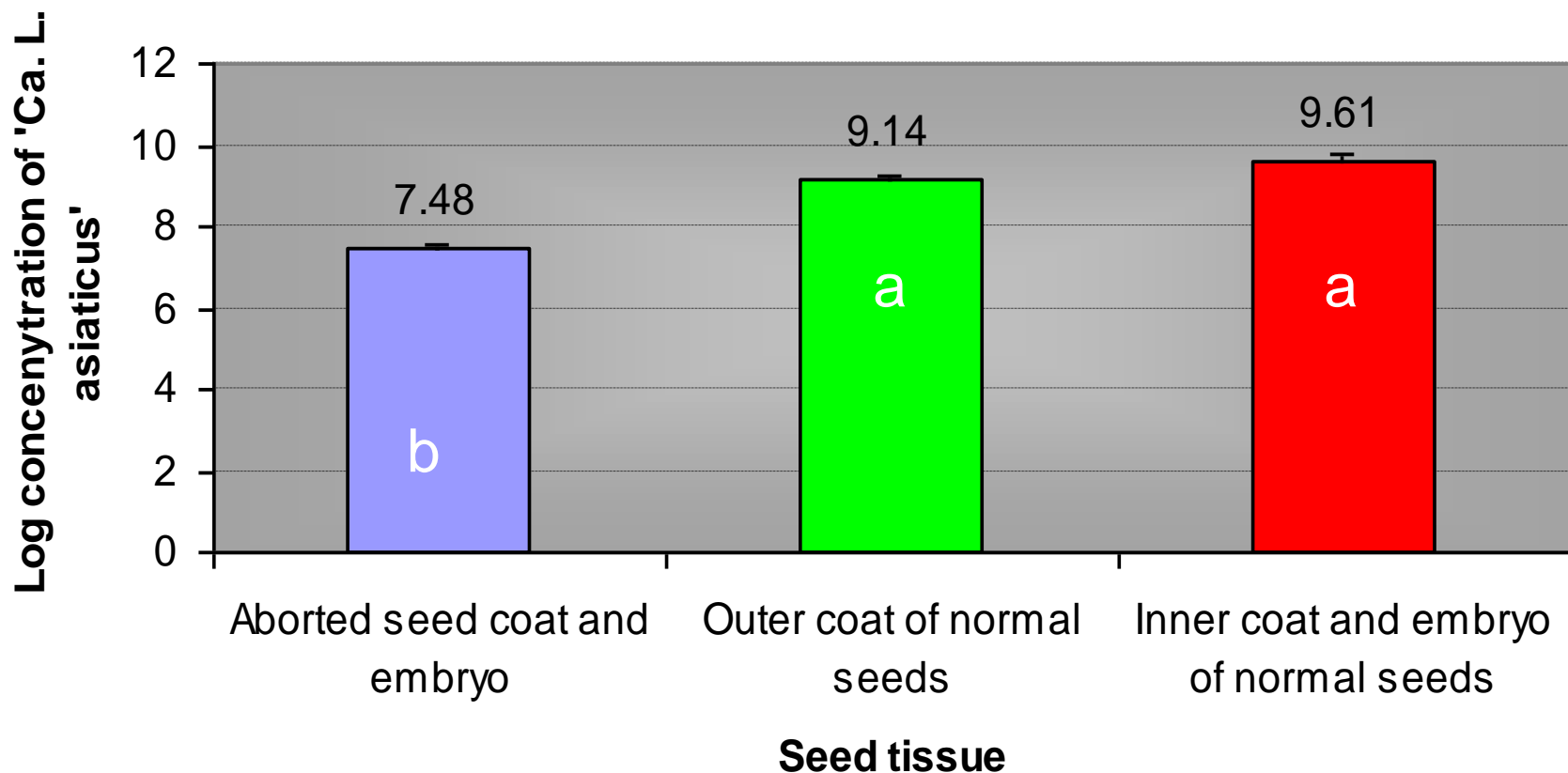


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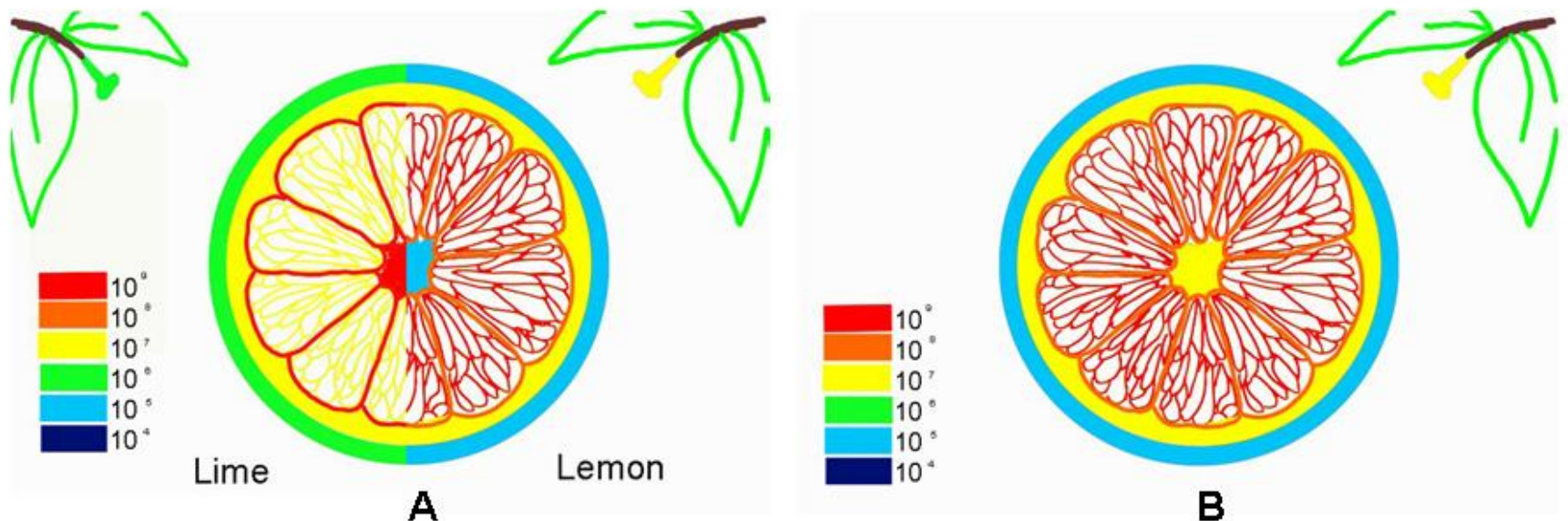
# Quantification of 'Ca. L. asiaticus' in seed tissues of HLB-affected *Citrus limonia* (lime)



Li 2005, unpublished



# Distribution of 'Ca. L. asiaticus' in fruit tissues of HLB-affected citrus species



**Figure. A.** Distribution of Ca Las genomes in fruit of lime and lemon. The colors indicate the relative concentration of Ca Las genome equivalents per gram of tissue sampled. The peduncles are drawn in the upper corners of the figure. **B.** Distribution of Ca Las genomes in fruit of lime and lemon averaged over the two fruits.

Li *et al.*, 2009. *Phytopathology* 99:139-144

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# Plant sample processing

- clean the samples
- get the midribs and petioles
- cut them into ~2 mm pieces



For Qiagen DNeasy mini kits, 100 to 200 mg per extraction.

For CTAB or modified CTAB, 200 to 500 mg per extraction.

Use a closed homogenization system, *i.e.* Lysing Matrix A tubes by a FastPrep or MiniBeadBeater machine

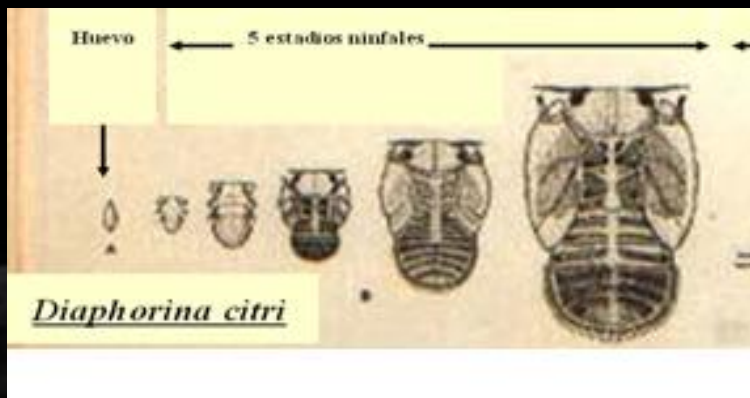


# Vector psyllids samples



If necessary, the psyllid samples can be stored at 4°C for more than one year for DNA extraction and PCR assays.

- Collect nymph and adult psyllids as many as possible on HLB symptomatic or suspect trees.
- Put the psyllids in individual bottles containing 70% ethanol (alcohol)

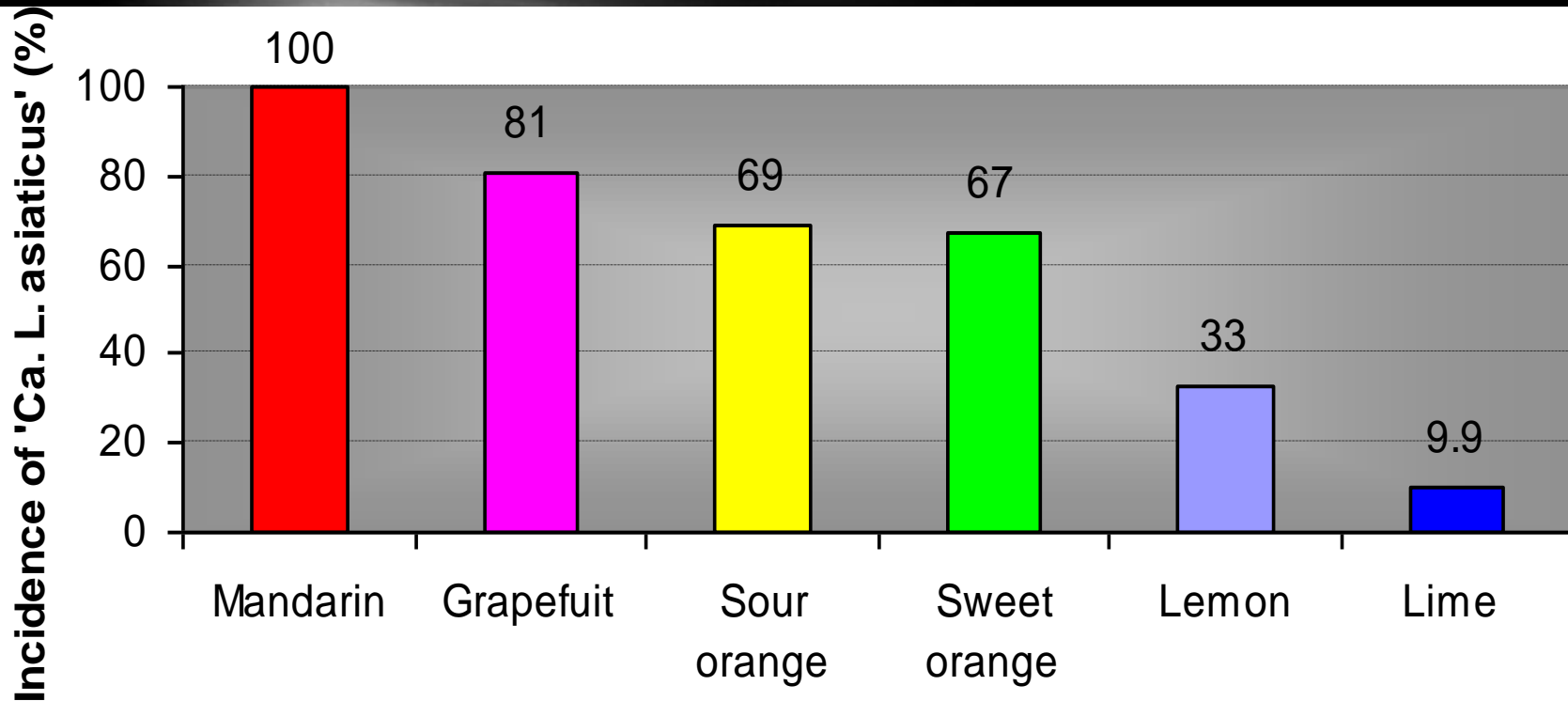


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# Incidence of 'Ca. L. asiaticus' in adults of *Diaphorina citri* collected on HLB symptomatic trees of citrus species in the field in Florida , Feb. to Mar. 2007



**Psyllids collected on HLB symptomatic trees of citrus species in the field in Florida**

Li et al., 2008. IRHLB p.231

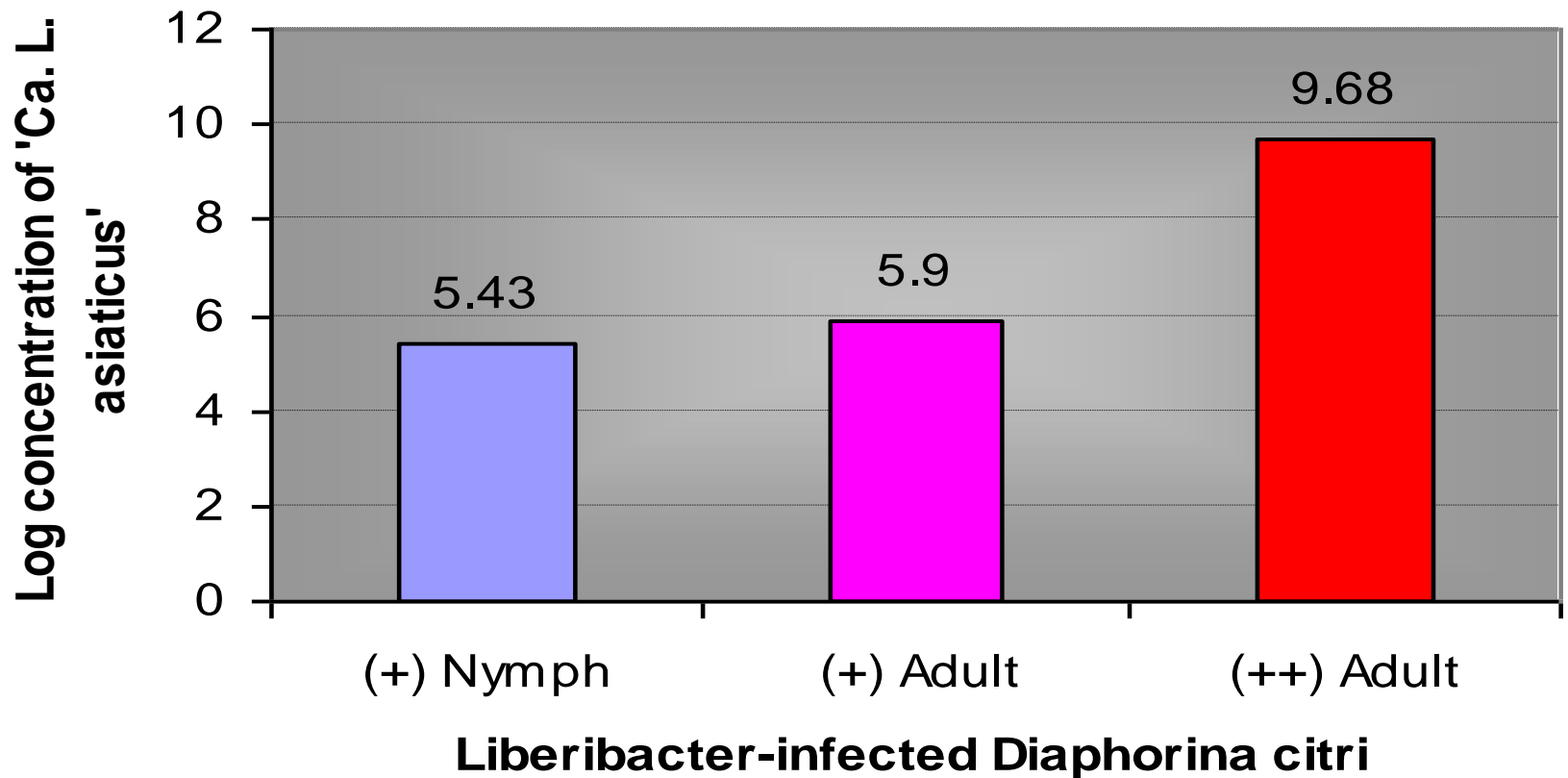


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Population of 'Ca. L. asiaticus' in adults and nymphs of *Diaphorina citri* collected on HLB symptomatic trees of citrus species in the field in Florida , Feb. to Mar. 2007



Li et al., 2008. IRHLB p.231



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# Insect sample processing



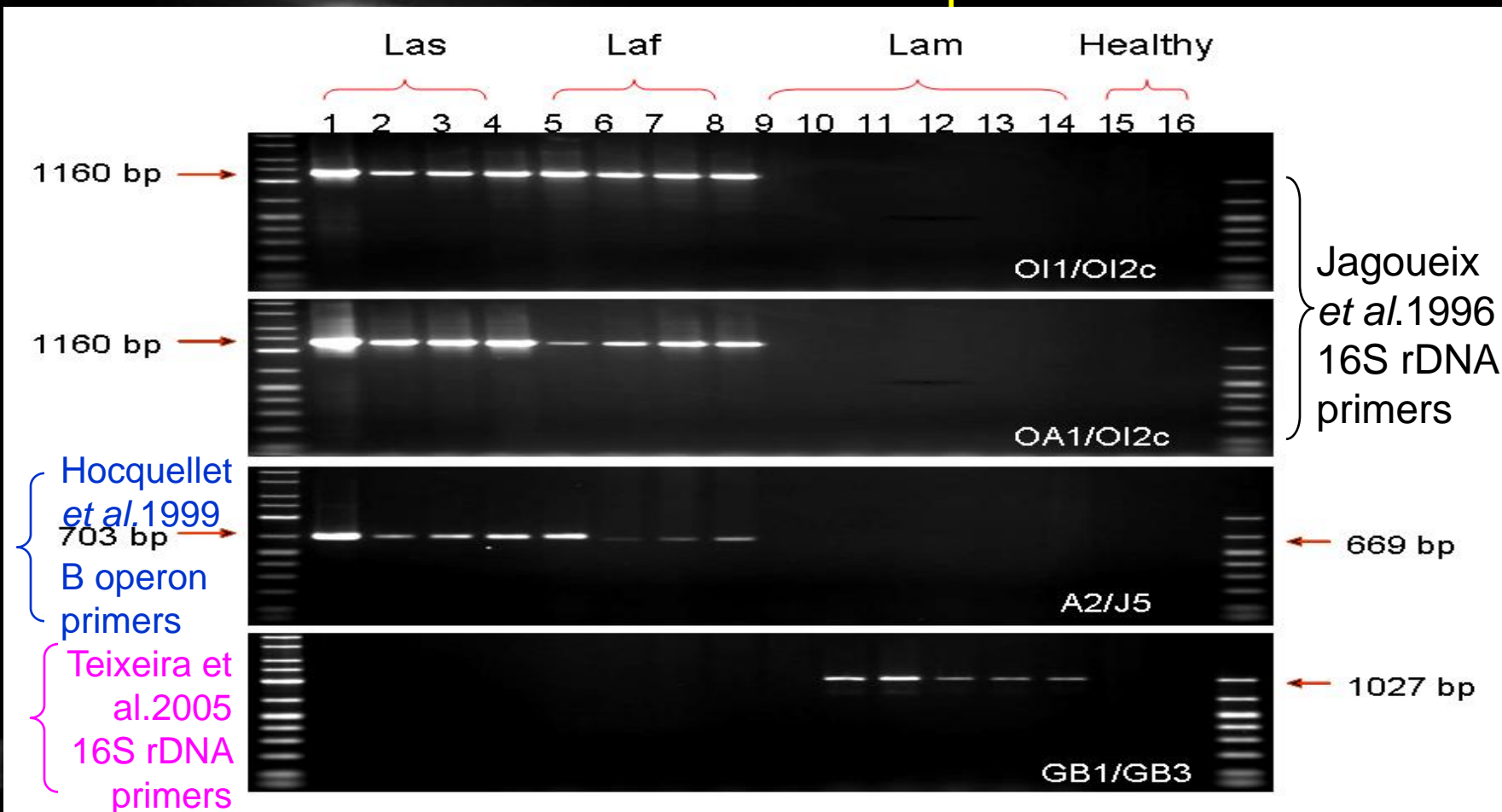
- Take psyllids from 70% alcohol on absorbent paper
- air dry the psyllids for 5-10 min
- take them to homogenization tubes or containers

For Qiagen DNeasy mini kits, 1 - 50 nymphs or 1 – 5 adults per extraction (USDA-APHIS).



Use a closed homogenization system, *i.e.* Lysing Matrix A tubes by a FastPrep or MiniBeadBeater machine

# Specificity of conventional PCR primers specific to 'Ca. Liberibacter sp.'



Source: Li et al., 2007. *Plant Dis.* 91:51-58.



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# Optimization and Standardization of Conventional PCR Protocol

## 1. Optimization of reagents.

Chemicals	Volume ( $\mu$ l)	Final concentration
Water	16.05	N/A
10x PCR buffer	2.5	1x
MgCl <sub>2</sub> (50 mM)	1.25	2.5 mM
dNTPs (10 mM each)	0.5	240 $\mu$ M each
Platinum Taq (5U/ ( $\mu$ l)	0.2	1 Unit
primers (2 $\mu$ M each)	2.5	200 nM
DNA sample	2.0	200 ng – 5 fg
Total		25 $\mu$ l

## 2. Standardization of the PCT protocol using a MJ Research PTC-200.

1 cycle	94° C for 2 min
	94° C for 30 S
35 cycles	62° C (OI1/2c and A2J5) or 54° C (GB1/3) for 1 min
	72° C for 1 min
1 cycle	72° C for 10 min

Source: Li et al.,2007. Plant Dis. 91:51-58.

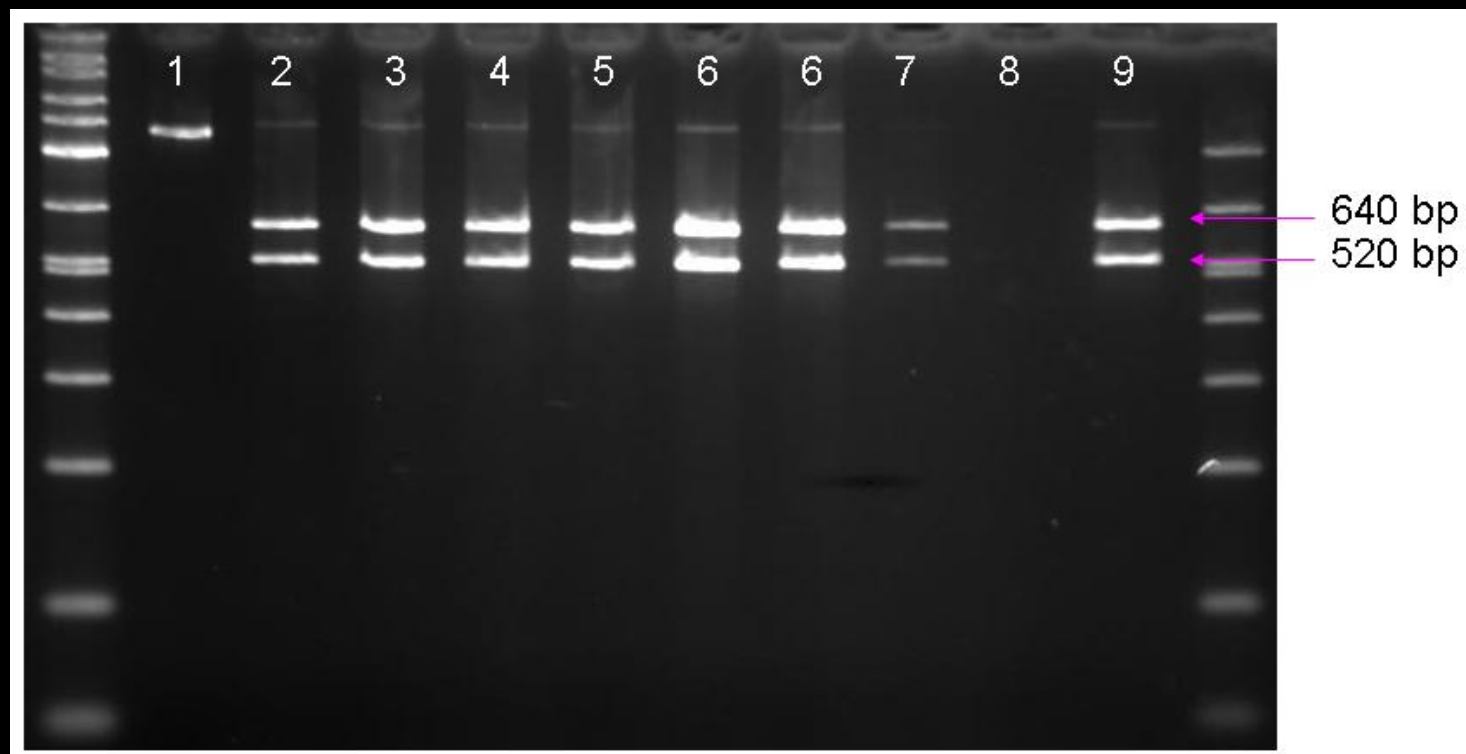


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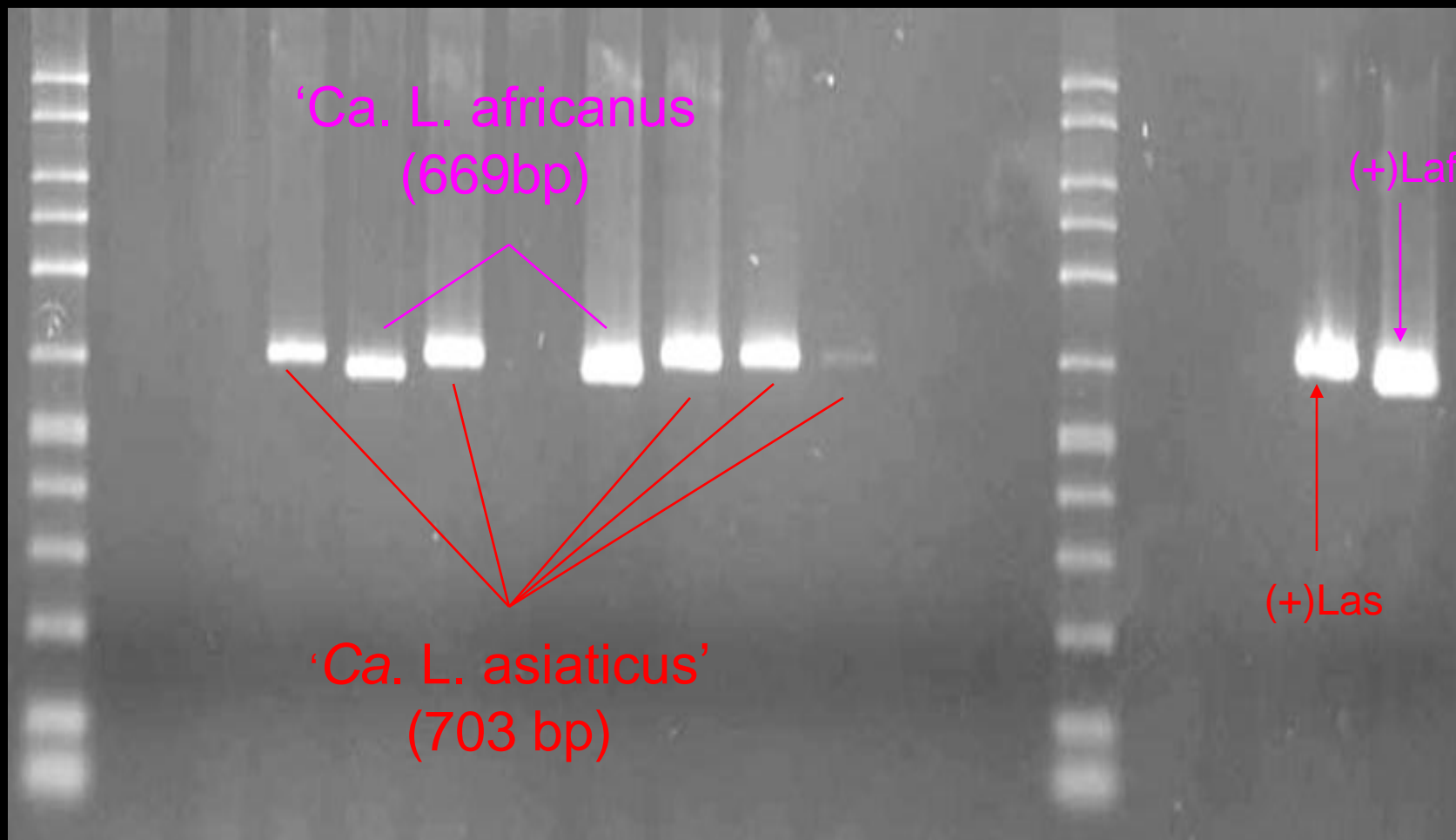
# Differentiation of 'Ca. L. asiaticus' from 'Ca. L. africanus' by Bové's 16S rDNA primers together with digestion by XbaI



**Figure.** Digestion (with Xba I) of PCR products obtained with O11/2c to differentiate Las from Laf. (Las: 640bp and 520bp; Laf: 520bp, 506bp, and 130bp)



# Differentiation of 'Ca. L. asiaticus' (Las) from 'Ca. L. africanus' (Laf) based on PCR amplicon sizes by Bové's $\beta$ operon primers



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# Optimization and Standardization of Real-Time PCR Protocol

## 1. Optimization of reagents.

Chemicals	Volume ( $\mu$ l)	Final concentration
Water	4.7	N/A
10x PCR buffer	2.5	1x
MgCl <sub>2</sub> (50 mM)	3.0	6.0 mM
dNTPs (10 mM each)	0.6	240 $\mu$ M each
Platinum Taq (5U/ ( $\mu$ l))	0.2	1 Unit
HLB primers (2 $\mu$ M each)	3.0	240 nM
HLB probe (1 $\mu$ M)	3.0	120 nM
COX primers (2 $\mu$ M each)	3.0	240 nM
COX probe (1 $\mu$ M)	3.0	120 nM
DNA sample	2.0	200 ng – 5 fg
Total		25 $\mu$ l

## 2. Standardization of the PCR protocol using a SmartCycler II.

Stage I	95° C for 20 s with optics off 40 cycles
Stage II	95° C for 1 s with optics off 58° C for 40 s with optics on

# Specificity of TaqMan PCR assays for '*Candidatus Liberibacter sp.*' in plant extracts

**Table. FAM Ct values of real-time PCR assays.**

Species, strain or isolate	Origin	HLBaspr	HLBampr	HLBasampr	
<i>Candidatus liberibacter asiaticus</i>	B239	Taiwan	22.51	0	22.26
	KIN1	Japan	25.26	0	24.89
	IDN5	Indonesia	26.32	0	25.99
	Br892	Brazil	21.61	0	21.05
	Fl136	Florida	20.36	0	24.25
<i>Candidatus liberibacter americanus</i>	Br974	Brazil	0	23.67	20.12
	Br875	Brazil	0	22.65	23.24
	Br976	Brazil	0	24.11	22.42
	Br977	Brazil	0	24.43	23.98
	Br978	Brazil	0	23.81	24.15
Las + Lam	Br892+974	Brazil	22.21	23.56	21.88
Other citrus pathogens or indophytes	Xac Strain A	Florida	0	0	0
	<i>X. fastidiosa</i> CVC strain	Brazil	0	0	0
	<i>P. Citricola</i> I 22F3	USA	0	0	0
	<i>P. citrophthora</i> I 1E4	USA	0	0	0
	<i>M. Mesophilicum</i> SR1.6/6	Brazil	0	0	0
	<i>C. Flaccumfacies</i> ER1/6	Brazil	0	0	0
	<i>P. Agglomerans</i> ARB18	Brazil	0	0	0
	<i>E. Cloacae</i> PR2/7	Brazil	0	0	0
	<i>Bacillus sp</i> CL16	Brazil	0	0	0
	DNA from plants with CTV T36	Florida	0	0	0
	DNA from plants with citrus blight	Florida	0	0	0
	DNA from plants with citrus sudden death	Brazil	0	0	0

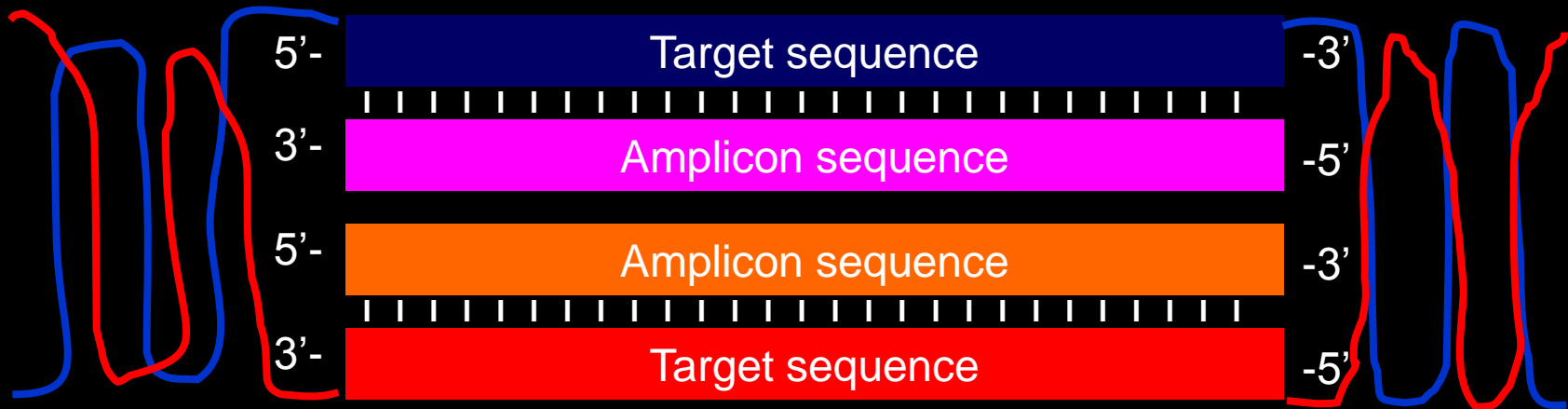
Source: Li et al., 2006. *J. Microbiol Methods* 66:104-115.

# PCR math

$$ACN = TCN \times 2^{\text{cycle}}$$

ACP: amplicon copy number (PCR products)

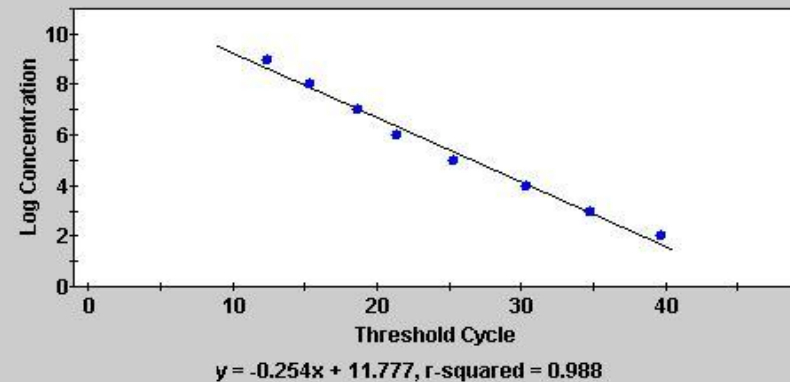
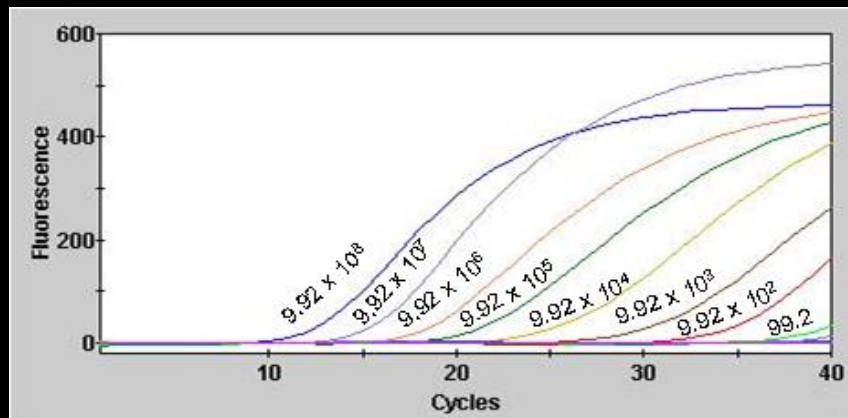
TCN: target copy number



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## Real-time PCR standard curve

$$Y = a + bX$$

Y: Log concentration of amplicon copy number

X: real-time Ct value (cycles)

a: Y interception

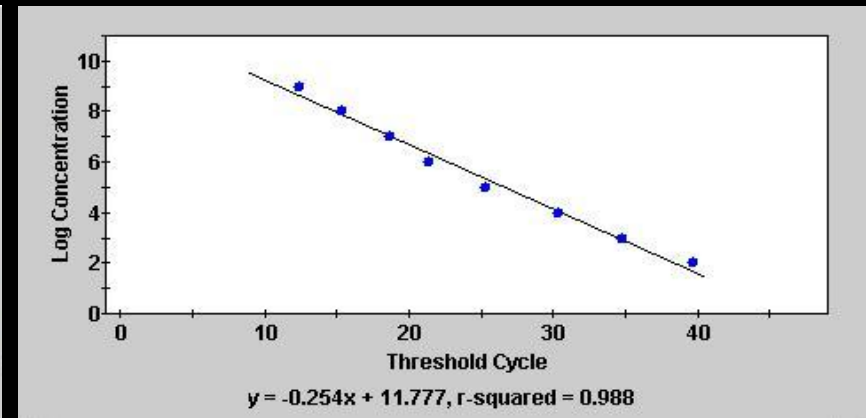
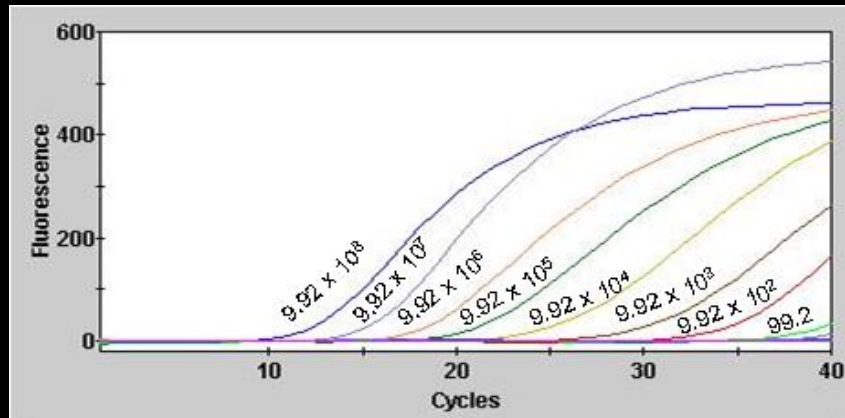
b: slope



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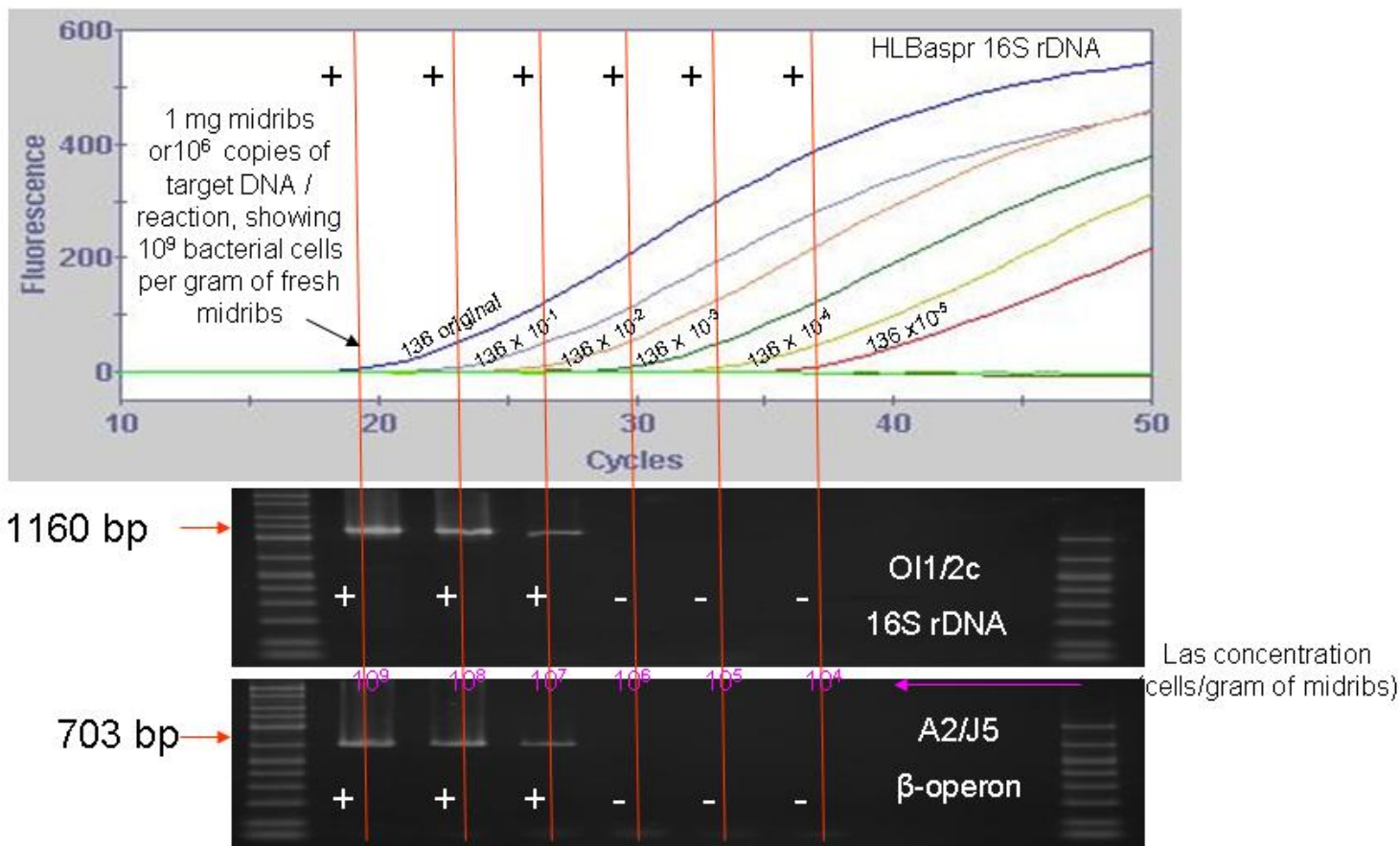


# Real-time PCR Amplification Efficiency

$$AE = 10^{-\text{slope}} - 1$$

slope: b of the real-time standard curve





**Figure.** Sensitivity comparison between real-time and conventional PCRs for a DNA sample (13600a) from a Las-infected tree of grapefruit in Florida.

Source: Li et al., 2007. *Plant Dis.* 91:51-58.



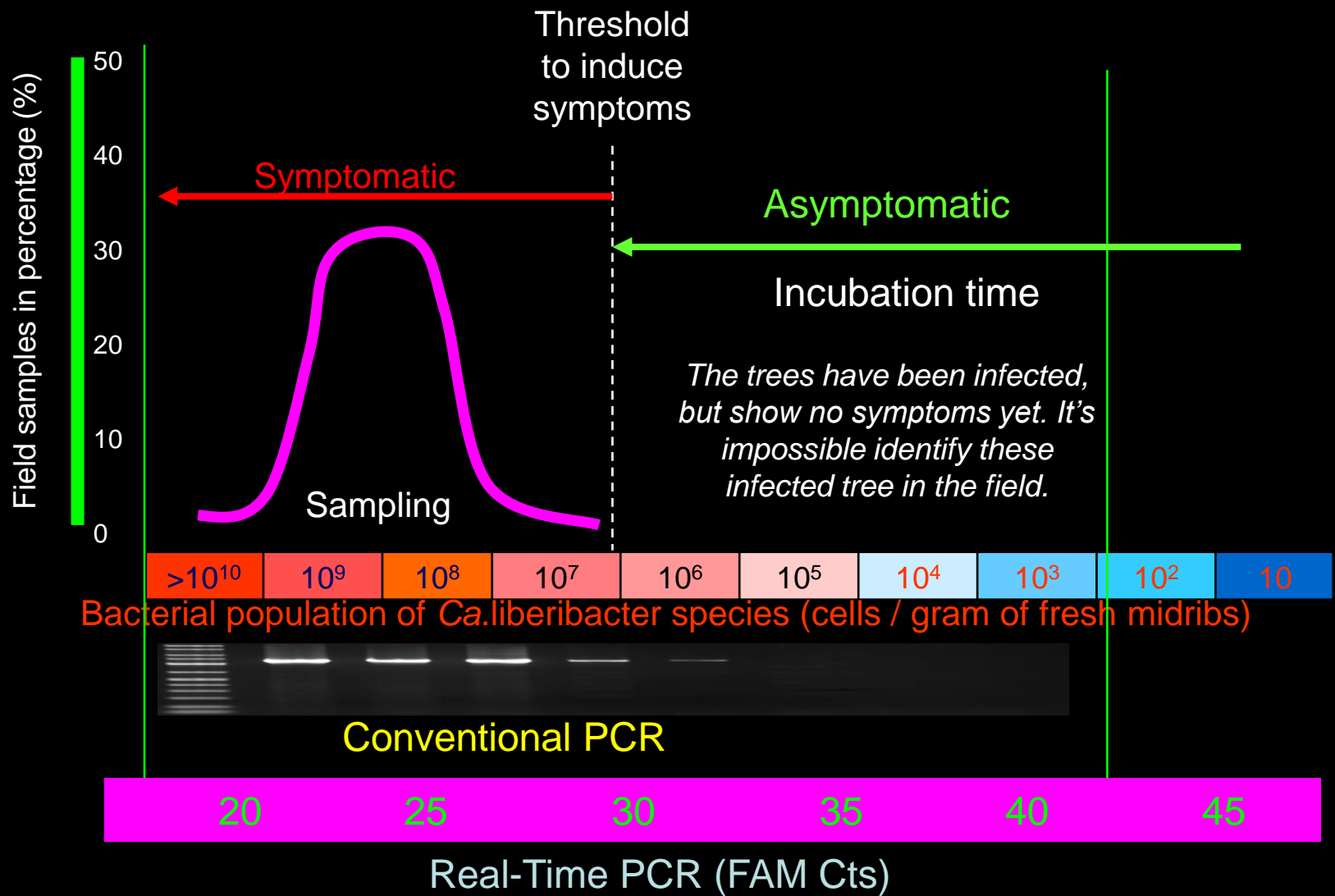
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**Table.** Real-time and conventional PCR for early detection of 'Ca. L. asiaticus' in citrus plants experimentally inoculated in a greenhouse.

Plant inoculated	Detection methods	Days post-inoculations			
		30	60	90	120
Common sweet orange	Symptoms	No	No	Yes	Yes
	Ct of qPCR	35.16	32.68	17.68	16.85
	cPCR-OI1/OI2c	-	-	+	+
	cPCR-A2/J5	-	-	+	+
Ridge pineapple sweet orange	Symptoms	No	No	No	Yes
	Ct of qPCR	0	34.24	30.35	19.49
	cPCR-OI1/OI2c	-	-	+	+
	cPCR-A2/J5	-	-	+	+
<i>C. hystrix</i>	Symptoms	No	No	No	Yes
	Ct of qPCR	0	35.06	31.32	20.14
	cPCR-OI1/OI2c	-	-	+	+
	cPCR-A2/J5	-	-	+	+



**Figure.** Comprehension of the Sensitivity of Real-Time PCR.



**Table.** Comparison of real-time and conventional PCR assays for detection of *Ca. L. asiaticus* Florida strain in single adult psyllids.

Sample	Real-time PCR		Conventional PCR		
	HLBaspr	WGfpr	OI1/OI2c	A2/J5	GB1/GB3
A1	25.34	27.55	+	+	-
A2	38.05	29.69	-	-	-
A3	0	28.34	-	-	-
A4	28.90	27.69	+	+	-
A5	27.08	27.98	+	+	-
B1	0	28.16	-	-	-
B2	0	28.92	-	-	-
B3	36.37	27.85	-	-	-
B4	31.39	28.13	+	+	-
B5	0	29.17	-	-	-
C1	0	29.08	-	-	-
C2	39.80	28.60	-	-	-
C3	0	29.06	-	-	-
C4	0	28.91	-	-	-
D1	0	28.23	-	-	-
D2	0	26.99	-	-	-
D3	0	28.09	-	-	-
D4	0	25.44	-	-	-
(+) ctrl	24.24	25.37	+	+	+
water	0	0	-	-	-



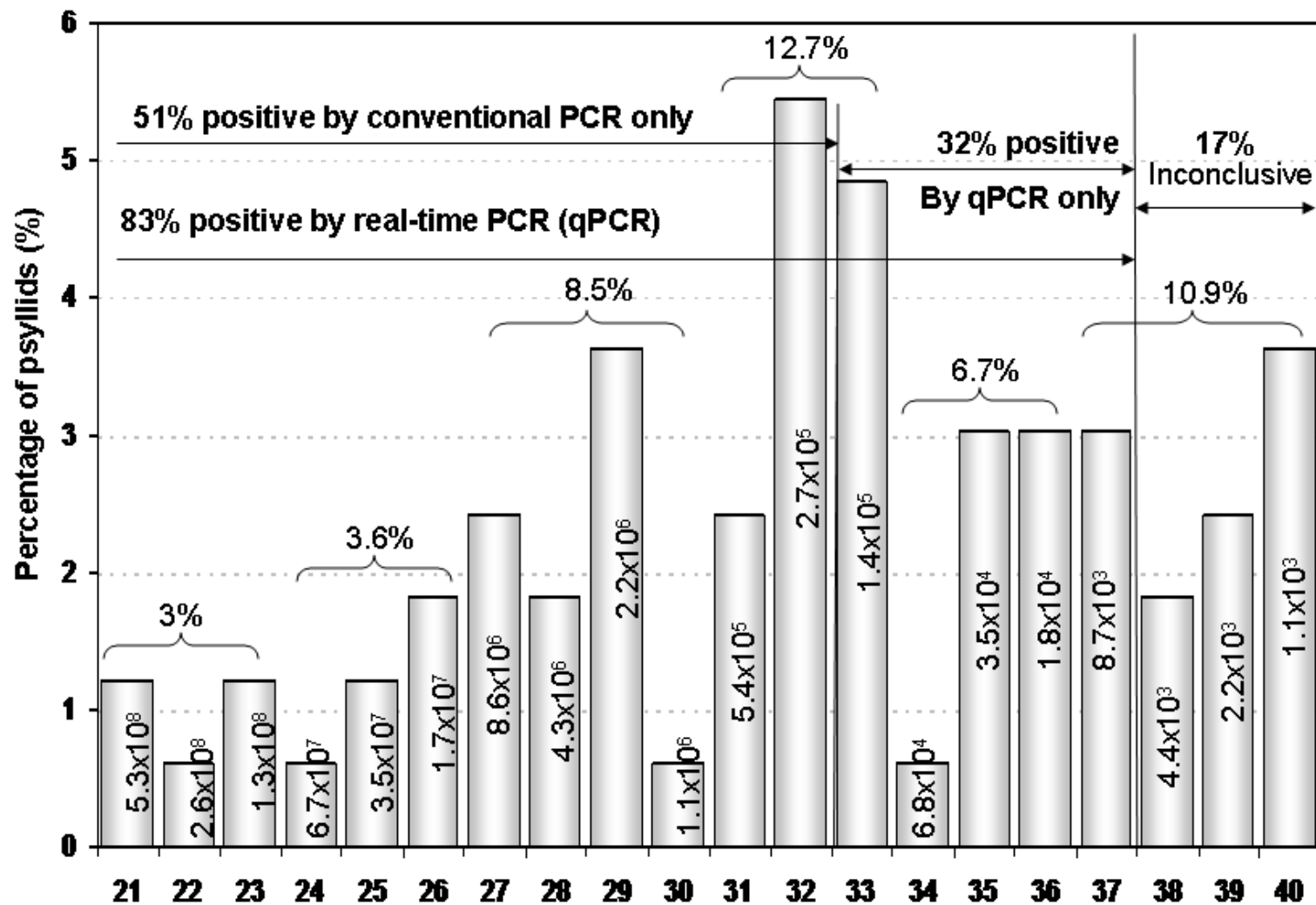
Source: Wenbin Li et al., 2008, Int Conf HLB, Orlando, FL



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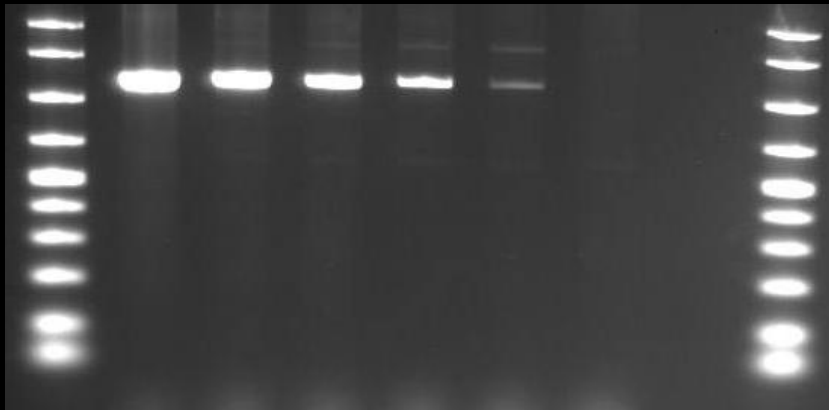


Ct Value of real-time PCR and *Ca. Liberibacter asiaticus* genomes (in column) per psyllid

Source: Wenbin Li et al., 2008, Int Conf HLB, Orlando, FL

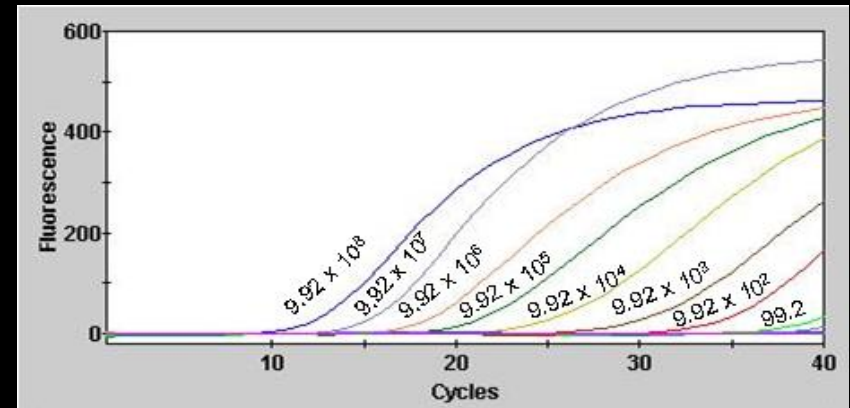
# Low Detection Limit (LDL)

The lowest target concentration (copy number) which can be detected at a probability of 95% and above by a detection method.



## Conventional PCR

1,000-10,000 target copies per reaction



## Real-time PCR

10-100 target copies per reaction

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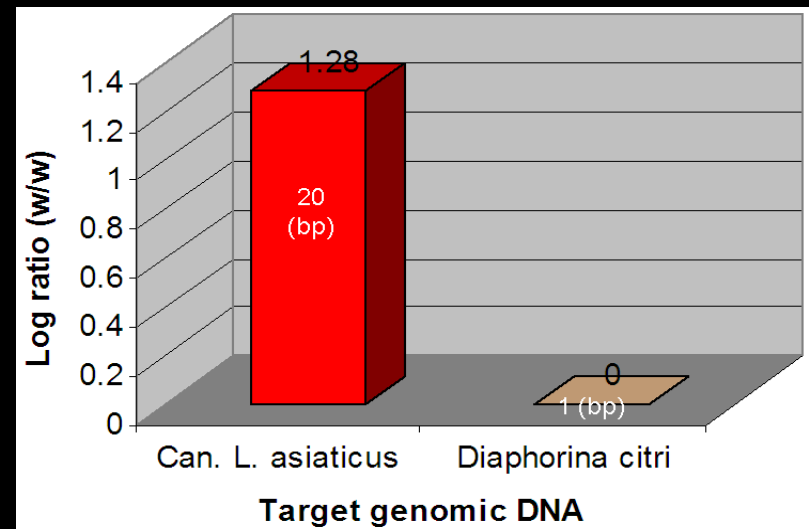
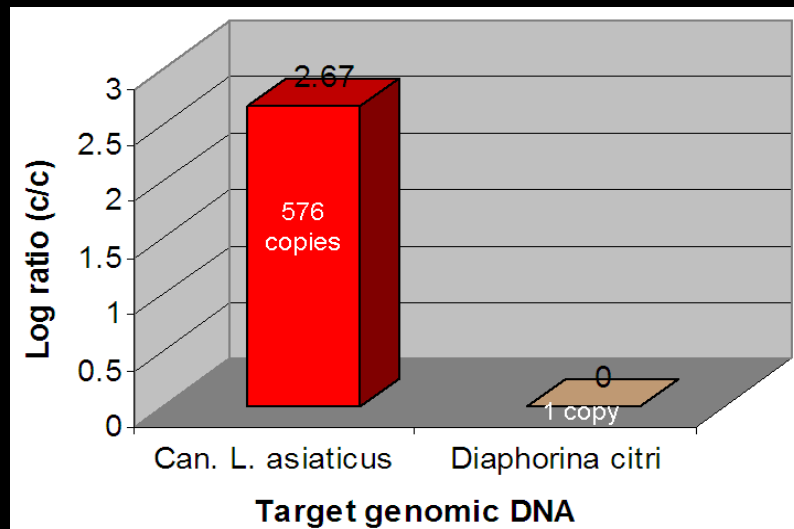
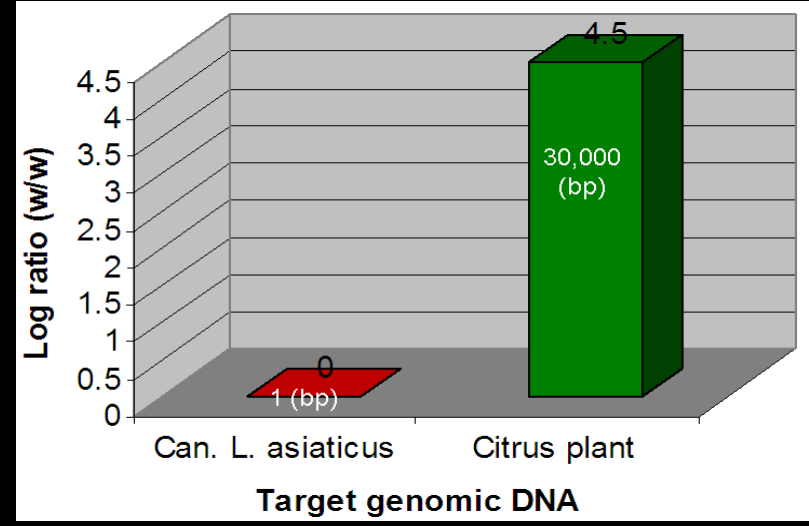
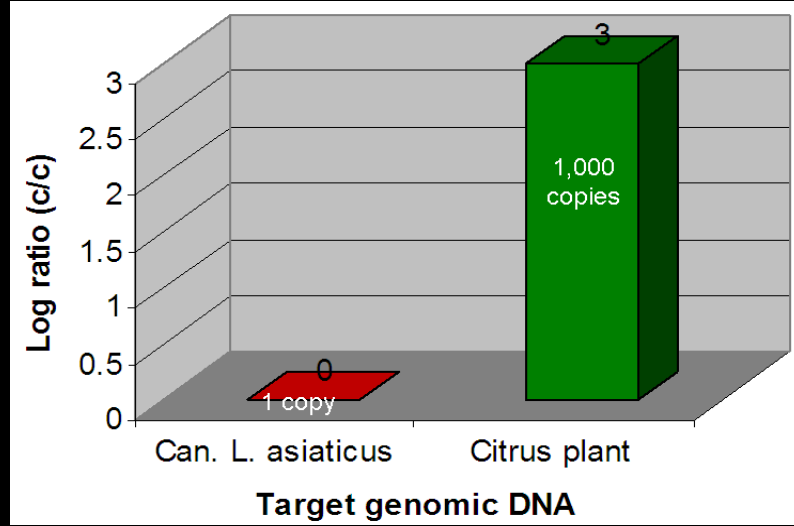


Figure. DNA ratios in extracts from Las-infected citrus and psyllids.

Source: Wenbin Li et al., 2006. JMM 66:104-115; 2008, Int Conf HLB, Orlando, FL





22:1011-1020, 2009

MPMI Vol. 22, No. 8, 2009, pp. 1011–1020. doi:10.1094/MPMI-22-8-1011.

e-Xtra\*

# Complete Genome Sequence of Citrus Huanglongbing Bacterium, '*Candidatus Liberibacter asiaticus*' Obtained Through Metagenomics

Yongping Duan,<sup>1</sup> Lijuan Zhou,<sup>2</sup> David G. Hall,<sup>1</sup> Wenbin Li,<sup>3</sup> Harshavardhan Doddapaneni,<sup>4</sup> Hong Lin,<sup>4</sup> Li Liu,<sup>5</sup> Cheryl M. Vahling,<sup>1</sup> Dean W. Gabriel,<sup>2</sup> Kelly P. Williams,<sup>6</sup> Allan Dickerman,<sup>6</sup> Yijun Sun,<sup>5</sup> and Tim Gottwald<sup>1</sup>

<sup>1</sup>USDA-ARS-USHRL, Fort Pierce, FL 34945, U.S.A.; <sup>2</sup>Dept. of Plant Pathology, University of Florida, Gainesville, FL 32653, U.S.A.; <sup>3</sup>USDA-APHIS-PPQ-CPHST, Beltsville, MD 20705, U.S.A.; <sup>4</sup>USDA-ARS, Parlier, CA 93648, U.S.A.; <sup>5</sup>ICBR, University of Florida, Gainesville, FL, U.S.A.; <sup>6</sup>Virginia Bioinformatics Institute Virginia Tech, Blacksburg, VA 24061, U.S.A.

Submitted 27 February 2009. Accepted 15 April 2009.



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In Press, 2009

## Confirmation of the sequence of '*Candidatus Liberibacter asiaticus*' and assessment of microbial diversity in Hunaglongbing-infected citrus phloem using metagenomic approach

Heather L. Tyler, Luiz W. Roesch, Siddarama Gowda, William O. Dawson and Eric W. Triplett

Dept Microbiology Cell Science, Dept Plant Pathology, CREC, University of Florida

- Used three next generation high-throughput sequencing platforms, 454, Solexa and SOLiD.
- Confirmed 99.99% of the sequence of '*Ca. L. asiaticus*' in HLB-infected citrus phloem.
- A culture independent, PCR-independent analysis of 16S rRNA sequences showed '*Ca. L. asiaticus*' was the only bacterium in HLB-infected citrus phloem.



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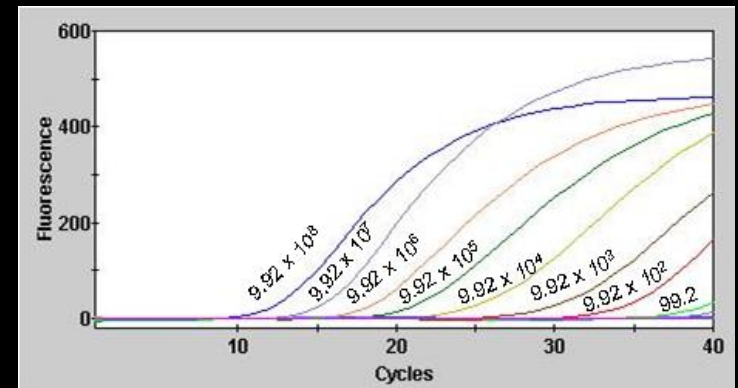
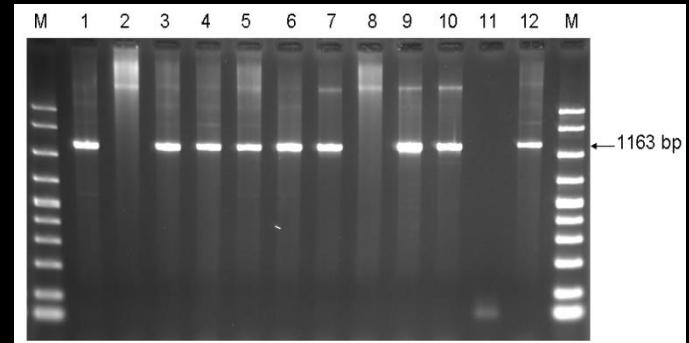


# Thank you!



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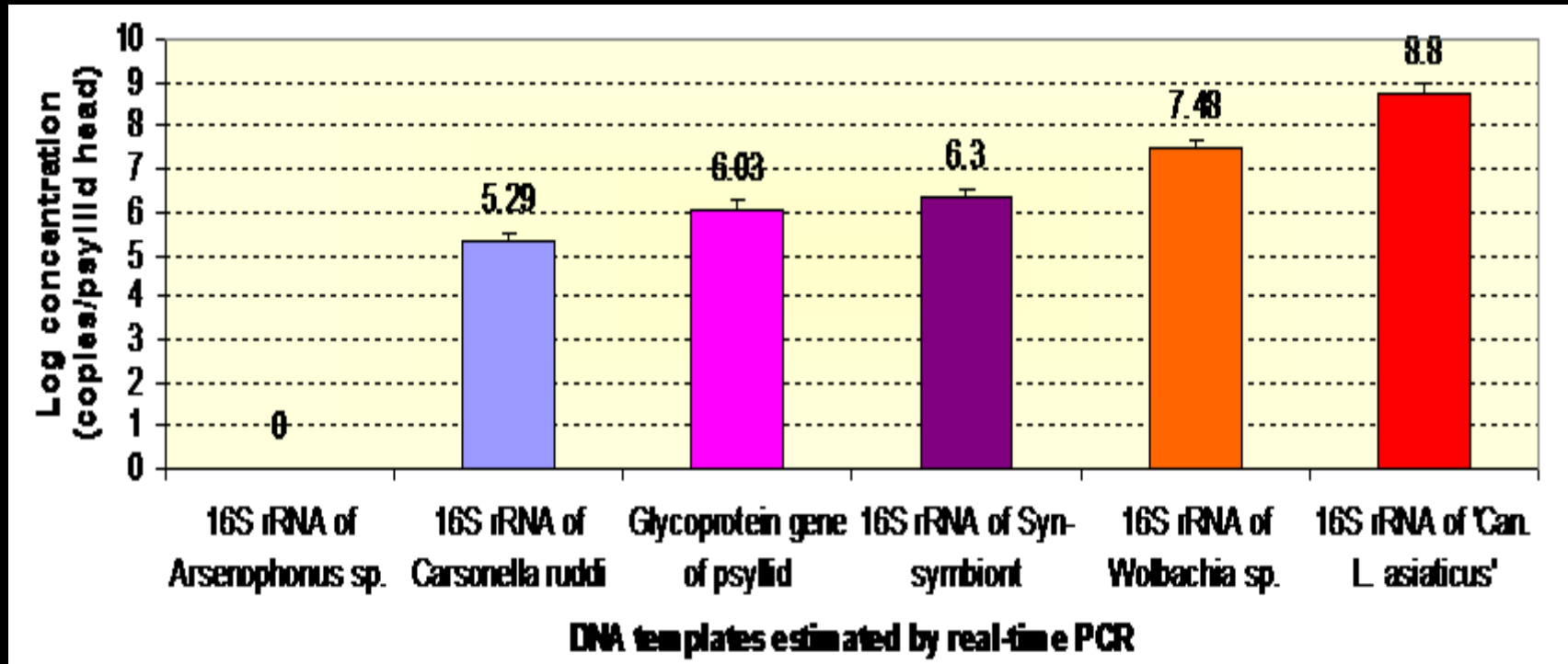




**Figure.** Zebra chip of potato and detection of the associated ‘*Ca. L. solanacearum*’ by real-time and conventional PCR.

Source: Li et al., 2009, *J Microbiol Methods* 78:59-65





**Figure.** Symbiont population in an adult of *Diaphorina citri* infected with '*Can. L. asiaticus*' in a citrus orchard in Florida.

Source: Wenbin Li et al., 2008, Int Conf HLB, Orlando, FL



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99: 480-485, 2009

## **Cultivation of ‘*Candidatus Liberibacter asiaticus*’, ‘*Ca. L. africanus*’, and ‘*Ca. L. americanus*’ Associated with Huanglongbing**

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Accepted for publication 13 December 2008.



United States Department of Agriculture  
Animal and Plant Health Inspection Service

**Plant Protection and Quarantine**

